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     (FILE 'HOME' ENTERED AT 09:58:14 ON 08 MAY 2002)

     FILE 'CAPLUS, EMBASE, BIOSIS' ENTERED AT 09:58:26 ON 08 MAY 2002
L1         17088 S REPROCESS?
L2       2098336 S TISSUE
L3         58565 S SLIDE OR SLIDES
L4           128 S L1 AND L2
L5             4 S L4 AND L3
L6             0 S THERMOSHANDON
L7             1 S "THERMO SHANDON"
L8        902140 S AUTOMAT? OR COMPUTER? OR MICROPROCESS?
L9             7 S L8 AND L4

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L4 ANSWER 67 OF 128 EMBASE COPYRIGHT 2002 ELSEVIER SCI. B.V.
AN 91226557 EMBASE
DN 1991226557
TI Method for **reprocessing** paraffin sections directly to resin sections for electron microscope microanalysis.
AU Blundell G.K.; Henderson W.J.
CS Biochemistry Department, University of Wales College of Cardiff, P.O. Box 98, Cardiff, South Glamorgan, United Kingdom
SO Journal of Histotechnology, (1991) 14/2 (109-111).
ISSN: 0147-8885 CODEN: JOHIDN
CY United States
DT Journal; Article
FS 001 Anatomy, Anthropology, Embryology and Histology
005 General Pathology and Pathological Anatomy
LA English

L4 ANSWER 59 OF 128 EMBASE COPYRIGHT 2002 ELSEVIER SCI. B.V.
AN 93339173 EMBASE
DN 1993339173
TI A method for detecting variability arising from errors in sample processing of paraffin-embedded **tissue** for DNA content analysis.
AU Hendricks J.B.; Hardt N.S.; Wilkinson E.J.; Pharis P.G.; Braylan R.C.
CS Department of Pathology, J. Hillis Miller Health Center, University of Florida DRL,Gainesville, FL 32610-0275, United States
SO Archives of Pathology and Laboratory Medicine, (1993) 117/11 (1138-1141). ISSN: 0003-9985 CODEN: ARPAAQ
CY United States
DT Journal; Article
FS 005 General Pathology and Pathological Anatomy
011 Otorhinolaryngology
LA English
SL English
AB We present a method for controlling variability that may arise from inconsistencies in sample preparation for DNA content analysis of paraffin- embedded **tissue**. Human tonsil **tissue** obtained from routine surgical specimens was embedded in paraffin according to standard protocols. Fifty-micrometer sections were cut from the block and analyzed each day for 20 days to establish control ranges. One tonsil **tissue** section was processed in parallel with each run of clinical specimens. In this context, a run was defined as the simultaneous processing of 50-.mu.m **tissue** sections for extraction of cell nuclei (dewaxing and rehydrating). If the tonsil G0/G1 peak coefficient of variation (CV) exceeded 2 SDs of the established mean, and optimum instrument performance and staining were verified, all samples prepared with the tonsil control were **reprocessed**. Instrument performance and staining were assessed by using the appropriate external controls. By using this rejection rule (1(2S)), the frequency of sample **reprocessing** in our laboratory was approximately 6%. When the run was repeated and the tonsil control CV was within acceptable range, the G0/G1 peak CV of the corresponding clinical specimens improved 25% of the time. Because most investigators are willing to accept higher CVs for paraffin-embedded **tissue** than for fresh **tissue**, it is desirable to have a control to detect decreased peak resolution, resulting from errors in sample processing.

L9 ANSWER 7 OF 7 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.
AN 1978:160351 BIOSIS
DN BA65:47351
TI THE CONTRIBUTION OF ELECTRON MICROSCOPY TO THE DIFFERENTIAL DIAGNOSIS OF TUMORS.
AU BONIKOS D S; BENSCH K G; KEMPSON R L
CS DEP. PATHOL., STANFORD UNIV. SCH. MED., STANFORD, CALIF. 94305, USA.
SO BEITR PATHOL, (1976) 158 (4), 417-444.
CODEN: BTPGAZ. ISSN: 0005-8165.
FS BA; OLD
LA English
AB From selected examples and results obtained by others, EM can, on occasion, be a significant aid in the accurate diagnosis of various neoplasms. While optimal fixation of tissues processed for EM produces excellent results with good cytologic preservation of the cellular organelles and extracellular components, the use of EM should not be excluded because the **tissue** is fixed in formalin or routinely embedded in paraffin for light microscopy. Rapid fixation of minced **tissue** in glutaraldehyde with postfixation in osmium tetroxide provides the best preservation of the fine structure of cells. Good results can be obtained if, instead of glutaraldehyde, minute pieces of **tissue** are originally fixed in buffered formaldehyde and processed according to standard EM techniques. Larger fragments of **tissue** fixed in formaldehyde and stored for prolonged periods of time can still be useful for diagnostic EM since many of the ultrastructural features used for diagnostic purposes are preserved even with prolonged formaldehyde fixation. Phosphate buffered commercial formaldehyde provides satisfactory fixation for routine light microscopy and EM. In the past many investigators suggested fixatives as substitutes for the usual light microscopy fixatives which could enhance ultrastructural preservation. All of these were financially unfeasible for large scale fixation or provided suboptimal fixation for light microscopy specimens. McDowell and Trump introduced the use of a phosphate buffered mixture of 4% commercial formaldehyde and 1% glutaraldehyde as a fixative which gives satisfactory preservation for routine **automated** histologic processing and EM studies. It should also be reemphasized that EM is possible not only in **tissue** fixed in formalin and processed shortly thereafter, but even when it is stored in formalin for prolonged periods of time, or **tissue** embedded in paraffin. A method of **reprocessing** for EM of formalin-fixed wet **tissue**, or formalin-fixed and paraffin embedded **tissue**, was described, as well as a method by which histologic sections of paraffin-embedded formalin-fixed postmortem specimens are prepared for EM study by what is called open-face embedding.